

Seco-Prezizaane-Type Sesquiterpenes and an Abietane-Type Diterpene from *Illicium minwanense*

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Received December 31, 2002

A methanol extract of the pericarps of *Illicium minwanense* afforded seven new *seco*-prezizaane-type sesquiterpenes (**2–8**) and a new abietane-type diterpene (**9**), together with six previously known compounds (**1** and **10–14**). The structures of the new compounds, (1*S**)- and (1*R**)-minwanenone (**2** and **3**), 1 α -hydroxy-6-deoxypseudoanisatin (**4**), (2*S*)-hydroxy-6-deoxypseudoanisatin (**5**), 3-oxopseudoanisatin (**6**), (3*S**,6*R**)-4,7-epoxy-6-deoxypseudoanisatin (**7**), 7-*O*-methylpseudoanisatin (**8**), and (+)-8,11,13,15-abietatetraene (**9**), were elucidated by spectroscopic data analysis and chemical transformations. The absolute configurations of **1** and **5** were established by X-ray crystallographic analysis of their *p*-bromobenzoyl derivatives.

Illicium species, which belong to the only genus of the family Illiciaceae, are widely distributed in southern regions of the People's Republic of China.¹ The cortex and root bark of *Illicium* species such as *I. merrillianum*, *I. jiadifengpi*, and *I. miwanense* occurring in China have been used in folk medicine for the treatment of rheumatoid arthritis. Their neurotoxicity, presumably due to anisatin,² however, sometimes limits their use in traditional medicine. *Illicium* species are rich in biosynthetically unique *seco*-prezizaane-type sesquiterpenes and prenylated C₆–C₃ compounds. Some *seco*-prezizaane-type sesquiterpenes have been found not to be neurotoxic, but to have an intriguing neurotrophic property unlike anisatin.^{3–6} In a preceding paper,⁷ we reported the structure and neurotrophic activity of jiadifenin, a new majucin-subtype of *seco*-prezizaane sesquiterpene isolated from *I. jiadifengpi*. As part of our ongoing studies on biologically active substances of the genus *Illicium*, we have investigated chemical components of the methanol extract of the pericarps of *I. minwanense*, which is indigenous to China, thereby resulting in the isolation of seven new *seco*-prezizaane-type sesquiterpenes (**2–8**) and a new abietane-type diterpene (**9**) along with the previously known compounds (**1** and **10–14**). In this paper, we report the structure elucidation of these new compounds and revise the previously proposed structure of miwanensin (**1**).⁸

Results and Discussion

A combination of silica gel and reversed-phase RP-8 column chromatography and preparative HPLC on the methanol extract of the pericarps of *I. minwanense* gave seven new *seco*-prezizaane-type sesquiterpenes and an abietane-type diterpene, (1*S**)- and (1*R**)-minwanenone (**2** and **3**), 1 α -hydroxy-6-deoxypseudoanisatin (**4**), (2*S*)-hydroxy-6-deoxypseudoanisatin (**5**), 3-oxopseudoanisatin (**6**), (3*S**,6*R**)-4,7-epoxy-6-deoxypseudoanisatin (**7**), 7-*O*-methylpseudoanisatin (**8**), and (+)-8,11,13,15-abietatetraene (**9**), along with the previously known compounds, miwanensin

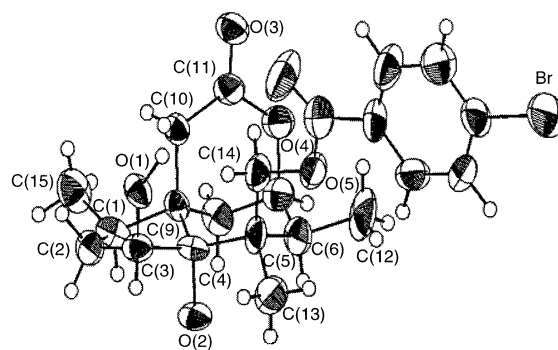


Figure 1. ORTEP drawing for **1a**.

(**1**),⁸ 6-deoxypseudoanisatin (**10**),⁹ pseudoanisatin (**12**),¹¹ 1 α -hydroxy-3-deoxypseudoanisatin (**13**),¹² and 1 α -hydroxypseudoanisatin (**14**).¹³

Miwanensin (**1**), a major component of *I. miwanense*, was reported with an α -oriented relative configuration of the hydroxyl group at the C-3 position, on the basis of comparing its NMR data with those of pseudoanisatin.¹⁰ However, the analysis of the NOESY data of **1** indicated that H-3 had no NOE interaction with H-10 β . Hence, we have decided to re-examine the absolute configuration of **1** by X-ray crystallography. Compound **1** was converted to its *p*-bromobenzoyl derivative **1a**, which gave a single crystal suitable for X-ray crystallographic analysis. Its ORTEP¹⁴ diagram is depicted in Figure 1 and indicates the absolute configuration for **1a**, in which the 3*R* configuration in **1** should be revised to 3*S*.

Compound **2** had a [M + H]⁺ ion peak at *m/z* 265.1411 in the high-resolution CIMS, corresponding to the molecular formula C₁₅H₂₀O₄, and its IR spectrum displayed absorptions due to a hydroxyl group at 3440 cm⁻¹, a lactone moiety at 1730 cm⁻¹, and a carbonyl group at 1705 cm⁻¹. The NMR spectral data (Table 1) of **2** contained signals corresponding to a minwanensin-type sesquiterpene except for signals due to H-10 β at δ_{H} 2.44 (q, *J* = 18.9, 2.6 Hz) and H-1 at δ_{H} 2.41 (q, *J* = 7.4 Hz) and the presence of an α,β -unsaturated ketone due to signals at δ_{H} 6.01 (s, H-3), δ_{C} 210.6 (C-2, CO), δ_{C} 131.3 (C-3), and δ_{C} 187.5 (C-4). The presence of an α,β -unsaturated ketone at the C-2, C-3, and

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Table 1. ^{13}C and ^1H NMR Spectral Data (δ) of Compounds **2**–**4**^a

position	2		3		4	
	δ_{C}	δ_{H}	δ_{C}	δ_{H}	δ_{C}	δ_{H}
1	53.8	2.41 q (7.4)	53.0	2.81 q (7.4)	82.9	
2 α	210.6		211.1		52.3	2.70 dd (15.5, 8.0)
2 β						1.93 dd (15.5, 8.0)
3	131.3	6.01 s	130.9	6.01 s	79.3	4.51 dd (8.0, 2.7)
4	187.5		188.2		83.8	
5	44.6		45.4		48.6	
6	46.1	1.94 qd (7.4, 2.4)	46.8	1.85 qd (7.4, 2.8)	49.4	2.95 q (6.9)
7	81.1	4.65 ddd (4.4, 2.4, 2.2)	81.3	4.68 ddd (3.8, 2.8, 2.2)	213.6	
8 α	37.5	1.87 ddd (13.7, 2.4, 2.2)	35.1	1.80 ddd (13.5, 2.2, 2.2)	42.4	3.00 dd (16.8, 1.8)
8 β		2.45 dd (13.7, 7.4)		2.31 dd (13.5, 3.8)		2.22 d (16.8)
9	45.8		46.2		51.5	
10 α	39.0	2.88 d (18.9)	41.7	2.89 d (19.0)	38.8	2.51 d (14.3)
10 β		2.44 dd (18.9, 2.6)		2.81 dd (19.0, 2.2)		3.57 dd (14.3, 1.8)
11	173.5		173.5		176.2	
12	12.4	1.20 d (7.4)	12.2	1.22 d (7.4)	8.2	1.10 d (6.9)
13	24.9	1.34 s	24.9	1.36 s	17.3	1.23 s
14 α	65.6	3.46 d (11.5)	65.4	3.48 d (11.5)	70.6	5.46 d (13.4)
14 β		3.54 d (11.5)		3.63 d (11.5)		3.78 d (13.4)
15	10.1	1.09 d (7.4)	10.1	1.10 d (7.7)	24.4	1.25 s

^a In CD_3OD at 600 MHz. J values (Hz) in parentheses.

C-4 positions was confirmed by HMBC correlations of H_3 -15 at δ_{H} 1.09 to C-1 at δ_{C} 53.8 and the C-2 carbonyl signal

(δ_{C} 210.6) and from H-3 at δ_{H} 6.10 to C-9 at δ_{C} 45.8. The relative stereochemistry on C-1 of **2** was elucidated by a NOESY experiment, in which CH_3 -15 and H-1 showed cross-peaks to H_2 -10 and H-8 α , respectively, indicating the H-1 β configuration. Thus, compound **2** was assigned as 2-oxo-3,4-dehydromiwanensin with a 1 S^* -configuration and has been named (1 S^*)-miwanenone.

Compound **3** was assigned the same molecular formula $\text{C}_{15}\text{H}_{26}\text{O}_4$ as **2**, obtained from high-resolution CIMS, and exhibited spectral data very similar to those of compound **2**. Additionally, the analysis of the 2D NMR data of **3** gave the same planar structure as **2**. This meant that **3** is a stereoisomer with respect to one of the chiral centers. In the NOESY experiment CH_3 -15 at δ_{H} 1.10 and H-1 at δ_{H} 2.81 showed NOE correlations to H-8 α and H_2 -10, respectively, but otherwise gave the same NOE enhancements as in **2**. Thus, the structure of **3** was assigned as (1 R^*)-miwanenone.

Compound **4** gave the molecular formula $\text{C}_{15}\text{H}_{22}\text{O}_6$, as determined by high-resolution FABMS at m/z 321 [$\text{M} + \text{Na}$]⁺, and its IR spectrum displayed absorptions due to a hydroxyl group at 3418 cm^{-1} and carbonyl groups at 1732 and 1716 cm^{-1} . The NMR spectral data of **4** (Table 1) were found to be similar to those of 6-deoxypseudoanisatin (**10**)⁹ except for the presence of the oxygenated quaternary carbon signal which resonated at δ_{C} 82.9 and the absence of any H-1 signal. The aforementioned spectral observations indicated that **4** has a hydroxyl group that replaces the proton on the C-1 position in **10**. Combined analysis of the ^1H - ^1H COSY, HMQC, and HMBC spectra of **4** led to the structure, 1-hydroxy-6-deoxypseudoanisatin. The relative configuration of the C-15 methyl group was assigned as β on the basis of the NOESY interaction between CH_3 -15 and H_2 -10. Thus, the structure of **4** was determined to be 1 α -hydroxy-6-deoxypseudoanisatin.

Compound **5** showed a molecular ion peak at m/z 298.1398 in the high-resolution EIMS, corresponding to the molecular formula $\text{C}_{15}\text{H}_{22}\text{O}_6$. The IR spectrum displayed the absorption attributable to a hydroxyl group at 3488 cm^{-1} and two carbonyl groups at 1735 and 1705 cm^{-1} . The ^1H and ^{13}C NMR spectral data (Table 2) of **5** showed signals similar to those of **10** except for the presence of two oxymethine signals at δ_{H} 4.32 (dd, $J = 7.4$ and 6.9 Hz) and δ_{C} 72.6 and at δ_{H} 4.25 (d, $J = 6.9$ Hz) and δ_{C} 78.5, which were assignable to C-2 and C-3 from the ^1H - ^1H COSY and

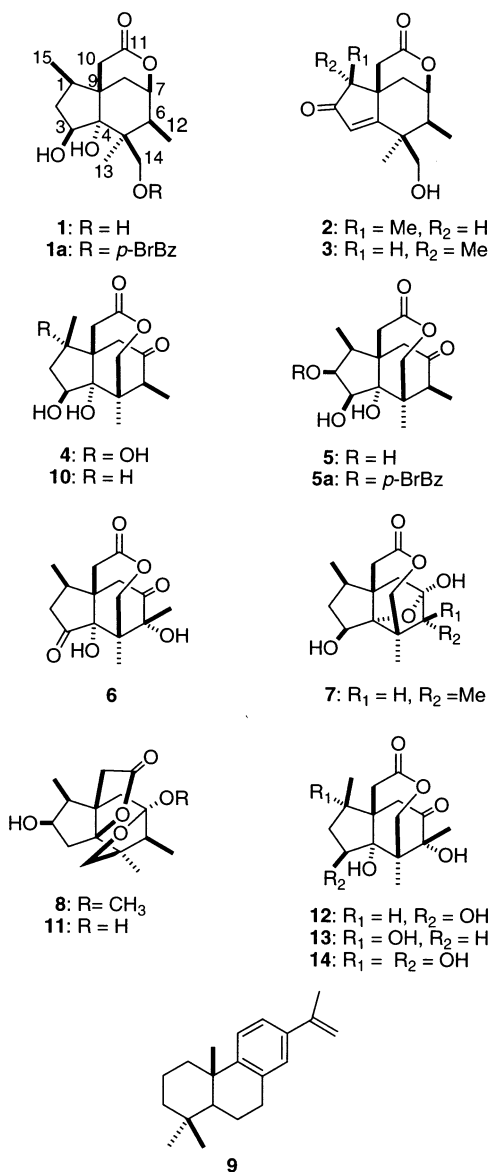
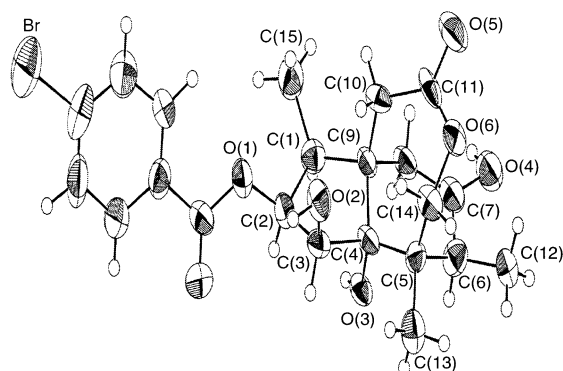


Table 2. ^{13}C and ^1H NMR Spectral Data (δ) of Compounds **5**^a and **6**^b

position	5		6	
	δ_{C}	δ_{H}	δ_{C}	δ_{H}
1	46.1	2.56 qd (7.4, 7.4)	35.9	3.01 qdd (19.0, 12.3, 7.1)
2 α	72.6	4.32 dd (7.4, 6.9)	41.5	2.82 dd (12.3, 9.3)
2 β				1.89 dd (19.0, 9.3)
3	78.5	4.25 d (6.9)	210.2	
4	82.4			78.8
5	48.6			46.4
6	49.7	2.88 q (6.9)	83.5	
7	213.3		205.2	
8 α	48.0	2.59 dd (16.2, 1.6)	40.8	2.83 d (15.8)
8 β		2.27 d (16.2)		3.38 dd (15.8, 2.2)
9	48.3		47.8	
10 α	36.7	2.31 d (14.8)	35.2	2.78d (14.9)
10 β		3.29 dd (14.8, 1.6)		2.84 dd (14.9, 2.2)
11	177.0		172.2	
12	8.1	1.10 d (6.9)	17.1	1.70 s
13	18.0	1.26 s	13.2	1.65 s
14 α	70.6	5.39 d (13.2)	69.7	4.60 d (14.0)
14 β		3.80 d (13.2)		4.08 d (14.0)
15	8.3	0.92 d (7.4)	13.0	0.82 d (7.1)

^a In CD_3OD at 600 MHz. ^b In $\text{C}_5\text{D}_5\text{N}$ at 600 MHz; J values (Hz) in parentheses.

**Figure 2.** ORTEP drawing for **5a**.

HMBC NMR spectra. These spectral data disclosed that **5** is 3-hydroxy-6-deoxypseudoanisatin. Treatment of **5** with *p*-bromobenzoyl chloride in pyridine and DMAP gave a single crystal of the *p*-bromobenzoyl derivative **5a**, for which X-ray crystallographic analysis was carried out. As a result, the absolute configuration on C-2 was determined to be *S*, as shown in the ORTEP¹⁵ diagram of **5a** (Figure 2). Thus, the structure of **5** was assigned as (2*S*)-hydroxy-6-deoxypseudoanisatin.

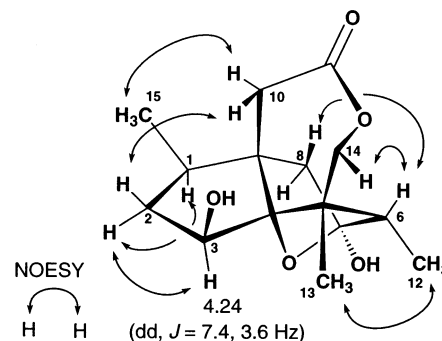
Compound **6** gave the molecular formula $\text{C}_{15}\text{H}_{20}\text{O}_6$, as determined by the high-resolution EIMS at m/z 296.1235, and its IR spectrum displayed absorptions due to a hydroxyl group at 3380 cm^{-1} and three carbonyl groups at 1737 , 1726 , and 1719 cm^{-1} . The NMR spectral data (Table 2) of **6** were similar to those of pseudoanisatin (**12**)¹¹ except for the absence of an oxymethine signal at C-3 existing in **12** and the appearance of a carbonyl signal that resonated at δ_{C} 210.2. These spectral data disclosed that the hydroxyl group on the C-3 position in **12** was oxidized to a ketone in **6**. The presence of the carbonyl group at this position was confirmed not only by the HMBC correlation of H-1 to the C-3 carbonyl signal but also a C-3 hydroxy group in pseudoanisatin (**12**) to **6** with Dess–Martin reagent. Thus, the structure of **6** was determined to be 3-oxopseudoanisatin.

Compound **7** was assigned the molecular formula $\text{C}_{15}\text{H}_{20}\text{O}_6$, as determined by the high-resolution EIMS at

Table 3. ^{13}C and ^1H NMR Spectral Data (δ) of Compounds **7**^a and **8**^b

position	7		8	
	δ_{C}	δ_{H}	δ_{C}	δ_{H}
1	45.1	2.33 qdd (12.6, 10.2, 7.1)	55.1	1.89 qd (7.2, 3.6)
2 α	45.0	2.53 ddd (13.8, 12.6, 7.4)	73.6	4.36 ddd (4.2, 3.6, 1.2)
2 β		1.36 ddd (13.8, 10.2, 3.6)		
3 α	73.8	4.24 dd (7.4, 3.6)	43.9	2.07 dd (15.0, 4.2)
3 β				2.33 dd (15.0, 1.2)
4	93.3		100.4	
5	40.5		50.9	
6	43.2	2.17 q (7.4)	44.1	1.86 q (7.2)
7	109.3		108.8	
8 α	47.9	1.56 d (12.5)	49.4	1.62 d (13.2)
8 β		1.73 d (12.5)		2.54 d (13.2)
9	52.9		48.6	
10 α	34.5	2.54 d (14.3)	41.6	2.78 d (18.6)
10 β		3.12 d (14.3)		3.20 d (18.6)
11	177.7		176.7	
12	10.4	0.92 d (7.4)	8.7	1.03 d (7.2)
13	16.1	1.13 s	14.0	0.91 s
14 α	72.2	4.83 d (13.1)	71.8	4.10 d (9.0)
14 β		3.79 d (13.1)		3.66 d (9.0)
15	14.1	0.94 d (7.1)	10.1	1.16 d (7.2)
OCH ₃			50.7	3.40 s

^a In CD_3OD at 600 MHz. ^b In $\text{C}_5\text{D}_5\text{N}$ at 600 MHz; J values (Hz) in parentheses.

**Figure 3.** Representative NOESY correlations of **7**.

m/z 282.1436, and its IR spectrum displayed absorptions due to a hydroxyl group at 3420 cm^{-1} and a carbonyl group at 1705 cm^{-1} . The NMR spectral data (Table 3) of **7** showed the presence of a tertiary methyl group (δ_{H} 1.13), two secondary methyl groups [δ_{H} 0.92 (d, $J = 7.4\text{ Hz}$), δ_{H} 0.94 (d, $J = 7.1\text{ Hz}$)], an oxymethylene [δ_{H} 3.79 and 4.83 (each d, $J = 7.1\text{ Hz}$)], and an oxymethine [δ_{H} 4.24 (dd, $J = 7.4, 3.6\text{ Hz}$); δ_{C} 73.8 (C-3)], which was coupled to a methylene [δ_{H} 2.53 (ddd, $J = 13.8, 12.6, 7.4\text{ Hz}$) and 1.36 (ddd, $J = 13.8, 10.2, 3.6\text{ Hz}$); δ_{C} 45.0 (C-2)]. The aforementioned spectral data indicated that **7** belongs to the 6-deoxypseudoanisatin-type sesquiterpenes as exemplified by **10**, but the presence of an acetal carbon signal at δ_{C} 109.3 instead of a carbonyl group on C-7 in **10** was the main difference in **7**. This acetal carbon showed HMBC correlations with both CH_3 -12 and H-8, and the chemical shift for C-4 in **7** appeared at δ_{C} 93.3. These spectral data, in addition to six degrees of unsaturation, indicated that **7** is a 4,7-acetal form of 6-deoxypseudoanisatin (**10**). However, compound **10** did not occur as an equilibrated mixture in CDCl_3 solution, whereas **7** was present as an acetal form in the same solution. The relative stereochemistry for **7** was elucidated on the basis of J values of vicinal coupling and NOESY data as shown in Figure 3. Thus, the large vicinal J value (7.4 Hz) between H-2 and H-3 indicated that

H-3 occurred in an α -configuration, and it was evident from the NOESY correlation of CH₃-15 to H₂-10 that the CH₃-15 was in a β -orientation. Additionally, the CH₃-12 signal showed a NOESY correlation with CH₃-13, and the H-6 signal showed NOESY correlations with both H-8 β and H-14, thereby indicating that CH₃-12 was in an α -configuration. Thus, compound **7** was elucidated as (3*S**,6*R**)-4,7-epoxy-6-deoxypseudoanisatin. It was not possible to determine why the stereochemistry on C-6 plays such a key role in eliciting a favorable equilibrium between the keto and acetal forms of pseudoanisatin on the basis of the global minimum energy calculated for **7** and **10** by MM2 molecular modeling.¹⁶

Compound **8** showed a protonated molecular ion peak at m/z 297.1714 [M + H]⁺ in the high-resolution FABMS, corresponding to a molecular formula of C₁₆H₂₄O₅. The IR spectrum displayed absorptions ascribable to a hydroxyl group (3441 cm⁻¹) and two carbonyl groups (1759 cm⁻¹). The ¹H and ¹³C NMR spectral data (Table 3) of **8** were identical to those of pseudomajucin (**11**)¹⁰ except for the presence of a methoxy group (δ_{H} 3.40; δ_{C} 50.7), and the chemical values of C-7 and C-8 were shifted to a lower field (δ_{C} 108.8) and a higher field (δ_{C} 49.4), respectively, compared with the corresponding data for **11**. This was taken as evidence that the C-7 hydroxy group in **11** is methylated. In fact, the methoxy signal showed a HMBC correlation to the C-7 acetal carbon at δ_{C} 108.8. Additionally, the NOESY experiment indicated that the relative stereochemistry for **8** was the same as that of **11**. To confirm the proposed structure of **8**, compound **11** was treated with orthotrimethylformate in the presence of Amberlyst-15 to give 7-*O*-methylpseudomajucin in 91% yield. Thus, the structure of **8** was established as 7-*O*-methylpseudomajucin.

Compound **9** showed in its high-resolution FABMS a peak at m/z 268.2198, corresponding to an elemental composition of C₂₀H₂₈. The IR spectrum displayed only the presence of a hydrocarbon. The ¹H and ¹³C NMR spectra of **9** were similar to those of (+)-8,11,13-abietatriene¹⁷ except for the presence of an olefin methyl group (δ_{H} 2.12, δ_{C} 21.8) and an *exo*-methylene [δ_{H} 5.01, 5.32 (each br s), δ_{C} 111.4 and 138.0]. The HMBC experiment displayed both the CH₃-16 and CH₂-17 signals correlated to C-13 at δ_{C} 143.1 and C-15 at δ_{C} 138.0; therefore the position of the *exo*-methylene group could be assigned to C-13. Thus, the structure of **9** was determined as (+)-8,11,13,15-abietatetraene.

In conclusion, we have isolated three miwanensin-type sesquiterpenes (**1–3**), eight pseudoanisatin-type sesquiterpenes (**4–7**, **10**, **12–14**), and two pseudomajucin-type sesquiterpenes (**8** and **11**) from *I. miwanense*. This plant is rich in pseudoanisatin-type sesquiterpenes, most of which have been isolated primarily from *I. ansitaum*.² According to the occurrence of these sesquiterpenes, *I. miwanense* is likely to be taxonomically close to the Japanese "Shikimi" (*I. ansitaum*). It is noted that none of the compounds isolated in the present study exhibited neurite outgrowth promoting activity on primary cultures of rat cortical neurons at 10 μM .¹⁸

Experimental Section

General Experimental Procedures. Optical rotations were measured on a JASCO DIP-1000 digital polarimeter. IR spectra were measured on a JASCO FT-IR 5300 infrared spectrophotometer. 1D- and 2D-NMR spectra were recorded on a Varian Unity 600 or 400 instrument. Chemical shifts are given as δ (ppm) with TMS as internal standard. MS were recorded on a JEOL AX-500 instrument. X-ray crystallography

was conducted on an MXC18 (MacScience Co.) instrument. Column chromatography was carried out on Keisegel 60 (70–230 mesh and 230–400 mesh).

Plant Material. The ripe fruits of *Illicium minwanense* were collected in Wenshang town, Yunnan, People's Republic of China, in September 1989, and a voucher specimen (No. 94041) is deposited at the Beijing University of Chinese Medicine.

Extraction and Isolation. The dried pericarps of *Illicium minwanense* (5 kg) were powdered and extracted with MeOH at room temperature to give 1.2 kg of a pale yellow extract. An aliquot of the MeOH extract (593.6 g) was divided into methanol-soluble (396.2 g) and methanol-insoluble portions (197.4 g). The methanol-soluble portion was chromatographed on a Celite column (515 g) eluting with a step gradient of *n*-hexane (100%), CH₂Cl₂ (100%), CH₂Cl₂–EtOAc (3:1 and 1:3), EtOAc (100%), and MeOH (100%) to yield six fractions (A–F).

Fraction A (15.2 g) was first subjected to Si gel chromatography eluting with *n*-hexane to give fractions 1–12. Fraction 2 (1.67 g) was further chromatographed on a Si gel column eluting with *n*-hexane giving fractions 13–19. Fraction 18 (7.5 mg) was separated by HPLC (Cosmosil Si 60, ϕ 10 \times 250 mm) using *n*-hexane as a solvent to give (+)-8,11,13,15-abietatetraene (**9**) (2.8 mg).

Fraction B (52 g) was chromatographed on a Si gel column eluting with a gradient of CHCl₃–EtOAc to give fractions 20–31. 6-Deoxypseudoanisatin (**10**) (134.8 mg), which was the major component, was obtained as crystals from fraction 25 (5.7 g). This fraction was chromatographed on Sephadex LH-20, with EtOH as the solvent, to give fractions 32–42. Fraction 36 (1.0 g) was further chromatographed eluting with CH₂Cl₂–MeOH (19:1) to give fractions 43–51. Finally, fraction 46 (13.7 mg) was purified by TLC (RP-18) with MeOH–H₂O–CH₃CN (1:6:3) to give 7-*O*-methylpseudomajucin (**8**) (5.3 mg). Fraction 25 (3.1 g) was chromatographed on a Lobar (RP-18) column eluting with MeOH–H₂O–CH₃CN (2:5:2) to give fractions 62–67. Fraction 62 (16.8 mg) was chromatographed on Sephadex LH-20, with EtOH as the solvent, to give pseudoanisatin (**12**) (11.5 mg). Fraction 64 (20.0 mg) was separated by reversed-phase HPLC (Cosmosil 5C₁₈-AR-II, ϕ 10 \times 250 mm) using MeOH–H₂O (3:2) to give (3*S**,6*R**)-4,7-epoxy-6-deoxypseudoanisatin (**7**) (2.8 mg). Fraction 30 (1.31 g) was chromatographed on a Si gel column eluting with CH₂Cl₂–EtOAc (1:5) and was purified by TLC with MeOH–H₂O (2:3) to give pseudomajucin (**11**) (179.9 mg).

Fraction 31 (1.41 g) was chromatographed on a Si gel column eluting with CHCl₃–EtOAc (1:5) to give fractions 68–75. Fraction 70 (171.2 mg) was chromatographed on a Lobar (RP-18) column eluting with MeOH–H₂O (1:1) to give minwanensin (**1**) (21.8 mg). Fraction 71 (187 mg) was further chromatographed on a Si gel column eluting with CHCl₃–MeOH (10:1) to give (1*R**)-minwanenone (**3**) (6.2 mg). Fraction 72 (142 mg) was further chromatographed on a Si gel column eluting with CHCl₃–MeOH (9:1) to give (1*S**)-minwanenone (**2**) (5.5 mg) and (2*S*)-hydroxy-6-deoxypseudoanisatin (**5**) (37.1 mg). Further TLC (RP-18) with MeOH–H₂O (2:3) of this fraction gave 1 α -hydroxy-6-deoxypseudoanisatin (**4**) (23.5 mg), 1 α -hydroxy-3-deoxypseudoanisatin (**13**) (13.7 mg), and 1 α -hydroxypseudoanisatin (**14**) (16.0 mg).

Fraction 49 (169.7 mg) was chromatographed on a Lobar (RP-18) column eluting with H₂O–CH₃CN (7:3) to give fractions 52–61. Fraction 56 (91.4 mg) was separated by reversed-phase HPLC (Cosmosil 5C₁₈-AR-II, ϕ 10 \times 250 mm) using MeOH–H₂O–CH₃CN (2:1:1) to give 3-oxopseudoanisatin (**6**) (1.0 mg).

***p*-Bromobenzoylation of Minwanensin (1).** To a solution of **1** (5.4 mg, 0.019 mmol) in pyridine (2 mL) was added *p*-bromobenzoyl chloride (41 mg) and 4-(dimethylamino)pyridine (4.2 mg). The reaction mixture was stirred at room temperature for 24 h and then condensed in vacuo to give a residue, which was purified by TLC with CHCl₃–MeOH (9:1) to afford **1a** (3.0 mg, 56%) as colorless plates; mp 229–230 °C. X-ray crystallographic analysis confirmed the structure of **1a** (absolute stereochemistry; ORTEP diagram, Figure 1).

Single-Crystal X-ray Crystallography of 1a.¹⁴ Suitable colorless plates of **1a** were obtained from a solution in CH₂-Cl₂; C₂₂H₂₈O₆Br. The crystal belongs to the monoclinic system, space group *P*2₁2₁2₁ with *a* = 8.440(3) Å, *b* = 11.265(6) Å, *c* = 27.668(2) Å, *V* = 2630.6(2) Å³, *Z* = 4, *D_x* = 1.468 Mg m⁻³, λ(Mo Kα) = 0.71073 Å. Intensity data were measured on a DIP image plate, θ_{max} = 25.73°. A total of 4004 reflections were collected. The structure was solved and refined by the *SHELXS-97* program.¹⁹ The non-hydrogen atoms were given anisotropic thermal parameters. The refinement converged to a final *R* = 0.0689, *R_w* = 0.1870 for 3297 observed reflections [*I* > 2.00σ(*I*)] and 293 variable parameters. The Flack parameter was 0.015(18). Values of the neutral atom scattering factors and real and imaginary dispersion corrections were taken from the *International Tables for X-ray Crystallography*.²⁰

(1S*)-Minwanenone (2): [α]_D¹⁹ -19.9° (*c* 1.25, MeOH); IR (film) ν_{max} 3440 (OH), 1730, and 1705 (C=O) cm⁻¹; ¹H and ¹³C NMR, see Table 1; HRCIMS *m/z* 265.1411 (calcd for C₁₅H₂₁O₄, 265.1440).

(1R*)-Minwanenone (3): [α]_D²¹ -17.9° (*c* 1.50, EtOH); IR (film) ν_{max} 3420 (OH), 1735, and 1683 (C=O) cm⁻¹; ¹H and ¹³C NMR, see Table 1; HRCIMS *m/z* 265.1434 (calcd for C₁₅H₂₁O₄, 265.1440).

1α-Hydroxy-6-deoxypseudoanisatin (4): [α]_D²¹ 7.6° (*c* 1.50, MeOH); IR (film) ν_{max} 3418 (OH), 1732, and 1716 (C=O) cm⁻¹; ¹H and ¹³C NMR, see Table 1; HRFABMS *m/z* 321.1343 (calcd for C₁₅H₂₂O₆Na, 321.1314).

(2S)-Hydroxy-6-deoxypseudoanisatin (5): mp 223–224 °C; [α]_D²⁰ -23.3° (*c* 0.38, MeOH); IR (film) ν_{max} 3488 (OH), 1735, and 1705 (C=O) cm⁻¹; ¹H and ¹³C NMR, see Table 2; HREIMS *m/z* 298.1398 (calcd for C₁₅H₂₂O₆, 298.1416).

***p*-Bromobenzoylation of (2S)-Hydroxy-6-deoxypseudoanisatin (5).** To a solution of **5** (2.3 mg, 0.0077 mmol) in pyridine (2 mL) was added *p*-bromobenzoyl chloride (16.9 mg) and 4-(dimethylamino)pyridine (3.8 mg). The reaction mixture was stirred at room temperature for 25 h and then condensed in vacuo to give a residue, which was purified by TLC with CHCl₃-MeOH (9:1) to afford **5a** (1.8 mg, 78%) as colorless plates; mp 214–215 °C. X-ray crystallographic analysis confirmed the structure of **5a** (absolute stereochemistry; ORTEP diagram, Figure 2).

Single-Crystal X-ray Crystallography of 5a.¹⁵ Suitable colorless plates of **5a** were obtained from a solution in CH₂-Cl₂; C₂₂H₂₆O₇Br. The crystal belongs to the monoclinic system, space group *P*2₁2₁2₁ with *a* = 6.72 Å, *b* = 13.04 Å, *c* = 28.07 Å, *V* = 2458.58 Å³, *Z* = 4, *D_x* = 1.530 Mg m⁻³, λ(Mo Kα) = 0.71073 Å. Intensity data were measured on a DIP image plate, θ_{max} = 25.67°. A total of 2145 reflections were collected. The structure was solved by the *maXus* program²¹ and refined by the *SHELXS-97* program.¹⁹ The non-hydrogen atoms were given anisotropic thermal parameters. The refinement converged to a final *R* = 0.063, *R_w* = 0.1590 for 1369 observed reflections [*I* > 2.00σ(*I*)] and 299 variable parameters. The Rogers η parameter was 1.044. Values of the neutral atom scattering factors and real and imaginary dispersion corrections were taken from the *International Tables for X-ray Crystallography*.²⁰

3-Oxopseudoanisatin (6): [α]_D²³ -181.2° (*c* 0.17, EtOH); IR (film) ν_{max} 3380 (OH), 1737, 1726, and 1711 (C=O) cm⁻¹; ¹H and ¹³C NMR, see Table 2; HREIMS *m/z* 296.1235 (calcd for C₁₅H₂₀O₆, 296.1260).

Dess–Martin Oxidation of Pseudoanisatin (12). To a solution of **12** (50.0 mg, 0.17 mmol) in CH₂Cl₂ (5 mL) was added Dess–Martin reagent ([1,1,1-triacetoxy-1,1-dihydro-1,2-benzodioxyl-3(1*H*)-one] periodinane) (62.2 mg). After being stirred at room temperature for 5 min, the solvent was evaporated in vacuo to give a residue, which was purified by TLC (RP-18) with MeOH-H₂O-CH₃CN (2:5:2) to afford 3-oxopseudoanisatin (**6**) (16.5 mg, 33%). This product was identical in all respects with natural 3-oxopseudoanisatin (**6**).

(3S*,6R*)-4,7-Epoxy-6-deoxypseudoanisatin (7): [α]_D²³ +42.6° (*c* 0.96, MeOH); IR (film) ν_{max} 3420 (OH), and 1705 (C=O) cm⁻¹; ¹H and ¹³C NMR, see Table 3; HREIMS *m/z* 282.1436 (calcd for C₁₅H₂₂O₅, 282.1467).

7-O-Methylpseudomajucin (8): [α]_D²³ -35.6° (*c* 1.08, MeOH); IR (film) ν_{max} 3441 (OH), and 1759 (C=O) cm⁻¹; ¹H and ¹³C NMR, see Table 3; HRFABMS *m/z* 297.1714 (calcd for C₁₅H₂₂O₅, 297.1702).

Methylation of Pseudomajucin (11). To a solution of **11** (2.0 mg, 0.007 mmol) was added orthotrimethylformate (250 μL) and Amberlyst-15 (three pieces). After being stirred at room temperature for 1 day, the solvent was evaporated in vacuo to give a residue, which was purified by TLC with CH₂-Cl₂-EtOAc (1:2) to afford 7-*O*-methylpseudomajucin (**8**) (2.0 mg, 91%). This product was identical in all respects with natural 7-*O*-methylpseudomajucin.

(+)-8,11,13,15-Abietatetraene (9): [α]_D²⁴ +48.2° (*c* 0.22, MeOH); IR (film) ν_{max} 1483 (C=C) cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 0.93 (3H, s), 0.97 (3H, s), 1.19 (3H, s), 1.46 (1H, br s), 1.75 (1H, m), 1.89 (1H, m), 2.12 (3H, s), 2.28 (1H, br s), 2.88 (1H, ddd, *J* = 16.4, 7.2, 1.7 Hz), 2.94 (1H, ddd, *J* = 16.4, 7.2, 1.7 Hz), 5.01 (1H, br s), 5.32 (1H, br s), 7.14 (1H, d, *J* = 1.7 Hz), 7.21 (1H, d, *J* = 8.0 Hz), 7.23 (1H, dd, *J* = 1.7, 8.0 Hz); ¹³C NMR (CDCl₃, 75 MHz) δ 38.8 (C-1), 19.1 (C-2), 41.7 (C-3), 33.5 (C-4), 50.4 (C-5), 19.3 (C-6), 30.5 (C-7), 135.0 (C-8), 149.6 (C-9), 37.7 (C-10), 124.3 (C-11), 122.9 (C-12), 143.1 (C-13), 126.1 (C-14), 138.0 (C-15), 21.8 (C-16), 111.4 (C-17), 33.3 (C-18), 21.7 (C-19), 24.8 (C-20); HREIMS *m/z* 268.2198 (calcd for C₂₀H₂₈, 268.2191).

Acknowledgment. We would like to thank Dr. Masami Tanaka and Miss Yasuko Okamoto for measuring the NMR and mass spectra. This work was partially supported by a Grant-in Aid for Scientific Research (No. 12480175) from the Ministry of Education, Culture, Sports, Science, and Technology of Japan.

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